Synthetic Studies on Spider Neurotoxins (I): Total Synthesis of Nephilatoxins (NPTX-9 and NPTX-11), New Neurotoxins of Joro Spider (*Nephila clavata*)

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Abstract: The first and efficient synthesis of Nephilatoxins (NPTX-9 and NPTX-11), new neurotoxins of Joro spider (*Nephila clavata*), has been achieved by employing a key azide intermediate as the polyamine unit.

Spider toxins such as NSTX-3, isolated from the Papua New Guinean spider (*Nephila maculata*),¹) and JSTX-3, isolated from the Joro spider (*Nephila clavata*),¹) have been demonstrated to be potent blockers of glutaminergic neuromuscular transmission in the mammalian synapse²) as well as invertebrates.³) Nephilatoxins (NPTX-1~12), new neurotoxins recently isolated from the Joro spider (*Nephila clavata*),⁴) have also been shown to have potent activities on the mast cell degranulation as well as specific interruption of glutaminergic neuromuscular transmission.^{4b} In particular NPTXs have been shown to have more potent histamine release activities from rat peritoneal mast cells than NSTX-3 and JSTX-3.^{4b} These toxins are structurally similar to NSTX and JSTX in that they possess a polycationic straight chain composed of basic amino acids and polyamines. However, in contrast to the latters, an indole-3-acetyl moiety instead of a 2,4-dihydroxyphenylacetyl moiety is connected at the N-termini of the polycationic chain of NPTXs, and the half of NPTXs contain ornithine as an amino acid unit which has not been found in other spider toxins.⁵)

Although spider toxins are rapidly emerging as unique tools for understanding excitatory amino acid transmission and related pharmacology, $^{2,3)}$ limited quantities have impeded their pharmacological evaluation and ongoing biological studies. We report here the first and efficient (practical) synthesis of NPTXs, NPTX-9 and NPTX-11, in which the azido group is employed as key functionality enabling the effective incorporation of the polyamine units. $^{6,7)}$

NPTX-9 and NPTX-11 have been assigned to the structures shown in Fig. 1,4) though the stereochemistry of amino acids has not been defined. NPTX-9, one of the most potent glutamate blockers among NPTXs,4b) consists of five components, i.e., an indole-3-acetyl-asparaginyl-cadaverine-ornithine-

arginine structure, while NPTX-11 is composed of the same structural units lacking of a terminating arginine residue of NPTX-9.





NPTX-9

The key point of the synthesis of spider toxins lies in the effective construction of the common polyamine units such as cadaverine (1,5-diaminopentane) and putreanine (8-amino-4-azaoctanoic acid) since each terminus of these α, ω -diamines is linked to a different amino acid, hence the use of the suitably masked α, ω -diamine or its equivalent is needed.⁸) In our synthetic strategy azide intermediates were designed as such ideally suited equivalents.

5-Azido-1-N-Boc-aminopentane (3), a cadaverine equivalent, was readily prepared starting from commercially available 5-amino-1-pentanol (1) in 96% overall yield in three steps⁹) (Scheme 1): 1) protection of the amino group with $(Boc)_2O$; 2) mesylation; 3) substitution with NaN₄.

With the cadaverine equivalent 3 in hand, we set out first to synthesize NPTX-11 by incorporating the L-forms of asparagine and ornithine.¹⁰⁾ Removal of a Boc group of 3 with trifluoroacetic acid (TFA) in CH₂Cl₂ followed by treatment with N-Boc-L-asparagine *p*-nitrophenyl ester in DMF containing triethylamine (TEA) gave 4 in 69% yield (Scheme 2). Similar treatment of 4 with TFA in CH₂Cl₂ followed by coupling with indole-3-acetic acid *p*-nitrophenyl ester in DMF in the presence of TEA furnished 5 having an indol-3-acetyl-asparaginyl-cadaverine structure in 76% yield. Both compounds 4 and 5 were easily purified by silica gel column chromatography, and the structures were confirmed by IR and ¹H-NMR. Catalytic hydrogenation of the azide 5 over 10% Pd-C in EtOH and subsequent coupling of the resulting amine with N^{\alpha}-Boc-N^{\delta}-Z-L-ornithine *p*-nitrophenyl ester in DMF gave the protected NPTX-11 (6) as amorphous solids in 89% yield. Removal of the Boc and the Z groups of 6 was readily accomplished under previously described conditions, i.e., TFA in CH₂Cl₂ followed by catalytic hydrogenation, yielding NPTX-11, which was purified by HPLC using a TOSOH TSK-gel ODS-120T column (water : acetonitrile : TFA= 85 : 15 : 0.1). The structure of the toxin, obtained in 51% yield, was confirmed by 270 MHz ¹H-NMR (D₂O), FD-MS 488 (M⁺), and HPLC.

On the other hand, treatment of 6 with TFA in CH_2Cl_2 followed by coupling of the resulting amine with tri-Z-L-arginine N-hydroxysuccinimide ester in DMF containing TEA yielded the fully protected NPTX-9 (7) in 60% yield (Scheme 2). The product could be purified by recrystallization from aqueous AcOH and the structure was confirmed by ¹H-NMR. Finally, deprotection of all the Z groups was readily accomplished by catalytic hydrogenation over 10% Pd-C in AcOH containing TFA to afford NPTX-9. The product was purified by HPLC using a JASCO Megapak SIL-C₁₈ column (gradient: A. 0.1% aq. TFA; B. 50% aq. acetonitrile containing 0.1% TFA) and obtained as TFA salt in 74% yield. All the data of the synthetic

Scheme 1



Reagents: a. (Boc)₂O, aq. Na₂CO₃, r. t. 12 h, 99%; b. MsCl, pyridine, CH₂Cl₂, O °C, 1 h; c. NaN₃, DMF, r. t. 12 h, 97% for the 2 steps.

Scheme 2



Reagents: d. TFA, CH₂Cl₂, r. t. 3 h; Boc-Asn-ONp, TEA, DMF, r. t. 12 h, 69%; e. TFA, CH₂Cl₂, r. t. 3 h; Indole-3-acetic acid *p*-nitrophenyl ester, TEA, DMF, r. t. 12 h, 76%; f. 10% Pd-C, H₂ (1 atm), EtOH, 3.5 h; Boc-Orn (Z)-ONp, TEA, DMF, r. t. 17 h, 89%; g. TFA, CH₂Cl₂, r. t. 3 h; 10% Pd-C, H₂ (1 atm), EtOH, 5 h, 51%; h. TFA, CH₂ Cl₂, r. t. 4 h; Z-Arg(Z₂)-ONSu, TEA, DMF, r. t. 12 h, 60%; i. 10% Pd-C, H₂ (1 atm), TFA, AcOH, 5 h, 74%.

compound (HPLC, 270 MHz ¹H-NMR, ¹³C-NMR, FD-MS 644 (M⁺), FAB-MS 645 (M+1)) was agreement with the proposed structure. The biological evaluation of the synthetic NPTX-9 and NPTX-11 by histamine release activity from rat peritoneal mast cells was also conformed to those of natural toxins.

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